 **Reproductive Genetic Innovations**

**2910 MacArthur Blvd, Northbrook, IL 60062**

**P: (847) 400-1515 F: (847) 400-1516**

[**info@rgiscience.com**](mailto:info@rgiscience.com) **www.RGIscience.com**

**PGT Dry Run Instructions**

*A dry run evaluates embryologists’ capability to place biopsy samples inside testing tubes properly for testing at RGI and to assess contamination levels.* ***No PGT results will be available on dry run samples and samples will be discarded after evaluation is complete.***

The collaborating IVF lab will receive at least 25 microcentrifuge tubes containing 5ml of lysis buffer for PCR. 10 biopsy samples are to be placed in ten separate tubes. Number each tube, 1 – 10. The medium surrounding each of the individual samples (i.e., micromanipulation drop) should be placed in individual tubes to be run as a control and labeled C1, C2, C3 …etc. Five tubes should remain unopened and marked NC for negative control.

All tubes are to be handled in the following manner:

1. Always handle tubes with gloves.
2. All tubes should be centrifuged in a microcentrifuge prior to loading the cell samples because they contain a small volume (5ml) of lysis buffer which may have been splashed along the side of the tube during handling.
3. Samples are transferred into tubes using a filtered stripper tip (inner diameter 120-135μm) under the control of a stereomicroscope while wearing gloves, a head cover and facemask. The volume of medium transferred with the sample should not exceed 1 microliter!
4. Stripper tips must be replaced between each sample in case of cell retention or lysis in the tip.
5. After, all the tubes containing samples are centrifuged once again and returned to the original transport box. The box is placed into the provided plastic bag as not to contaminate samples and then into a freezer (-20° C) until ready to ship.
6. A Dry Run Worksheet, detailing the contents of each tube, should accompany the specimens.

A dry run is considered successful if the evaluated sample amplification is 75% or higher and the contamination rate is less than 20%. Any result outside of this range will require a repeat of the dry run.

**If you have any questions, please feel free to call Yuri Ilkevitch, PhD., Director of Micromanipulation at 847-571-1392 or email** [**ivflab@rgiscience.com.**](mailto:ivflab@rgiscience.com.)

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**Dry Run Worksheet**

|  |  |  |
| --- | --- | --- |
| **Biopsy performed by:**  **\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_\_**  **(Last name, First Name)** | **Patient Name:**  N/A **D.O.B.:** N/A  **Type of Testing:** Dry Run  **Biopsies:** TE | |
| **IVF Center:** | **Date of Aspiration:**  N/A | **Date of Biopsy:**  N/A |
| **Labeling Legend:** Embryo # on top of the tube **NC**- Negative Control | | |
|  | | |

|  |  |  |
| --- | --- | --- |
| **Embryo #** | **Embryo Grade** | **# Of Cells Removed** |
| 1 |  |  |
| 2 |  |  |
| 3 |  |  |
| 4 |  |  |
| 5 |  |  |
| 6 |  |  |
| 7 |  |  |
| 8 |  |  |
| 9 |  |  |
| 10 |  |  |
| 1 NC |  |  |
| 2 NC |  |  |
| 3 NC |  |  |
| 4 NC |  |  |
| 5 NC |  |  |
| 6 NC |  |  |
| 7 NC |  |  |
| 8 NC |  |  |
| 9 NC |  |  |
| 10 NC |  |  |

**No PGT results will be available on dry run samples and samples will be discarded after evaluation is complete.**

***Please complete a separate worksheet for each participating embryologist.***

***Return samples to: RGI, 2910 MacArthur Blvd, Northbrook, IL 60062***